



Which Antibiotic Should We Use in Children Who Have had Hematopoietic Stem Cell Transplantation and Have MultiDrug Resistant Gram-Negative Infections? Single-Center Experience and Literature Review

Hematopoietik Kök Hücre Nakli Yapılmış ve Çoklu İlaç Dirençli Gram-Negatif Enfeksiyonları Olan Çocuklarda Hangi Antibiyotik Kullanmalıyız? Tek Merkez Deneyimi ve Literatür Taraması

Barış Malbora¹(ID), Hakan Sarbay²(ID), Dilek Ece²(ID), Derya Bayırlı Turan³(ID), Zeynep Doğusan⁴(ID), Abdullah Avni Atay²(ID)

¹ Unit of Pediatric Stem Cell Transplant, Yeni Yüzyıl Üniversitesi Gaziosmanpaşa Hospital, İstanbul, Türkiye

² Clinic of Pediatric Hematology and Oncology, Yeni Yüzyıl Üniversitesi Gaziosmanpaşa Hospital, İstanbul, Türkiye

³ Clinic of Infectious Diseases and Clinical Microbiology, Yeni Yüzyıl Üniversitesi Gaziosmanpaşa Hospital, İstanbul, Türkiye

⁴ Unit of Cell Processing, Yeni Yüzyıl Üniversitesi Gaziosmanpaşa Hospital, İstanbul, Türkiye

Cite this article as: Malbora B, Sarbay H, Ece D, Bayırlı Turan D, Doğusan Z, Atay AA. Which antibiotic should we use in children who have had hematopoietic stem cell transplantation and have multidrug resistant gram-negative infections? Single-center experience and literature review. J Pediatr Inf 2026;20(1):e62-e70.

Abstract

Multidrug resistant gram-negative bacteria infections are major problem especially in immunocompromised patients who have received hematopoietic stem cell transplantation. In this study, we aimed to compare the effects of colistin-based and ceftazidim-avibactam based therapy in children with hematopoietic stem cell transplantation. The medical records of pediatric patients who received hematopoietic stem cell transplantation between February 2019 and May 2021 were assessed retrospectively. A pre-structured case report form was used to record demographics, underlying diseases, severity of illness upon admission, detailed description of index infection requiring ceftazidim-avibactam administration, data related to colistin, ceftazidim-avibactam, or other antibacterial administration, clinical and microbiologic responses, outcome, and adverse events. A total of 183 pediatric patients had hematopoietic stem cell transplantation between February 2019 and May 2021. Eighty-nine bacterial infections were detected, and seventy-two Gram-negative microorganisms were isolated in 57 of the patients. Fourteen (24.6%) of these patients had 18 (25%) multidrug resistant gram-negative microorganisms. Nephrotoxicity occurred in 5 of 7 (71%) of the patients who received colistin-based treatment, while

Öz

Çoklu ilaç dirençli gram-negatif bakteri enfeksiyonları, özellikle hematopoetik kök hücre nakli yapılan hastalar gibi immün yetmezlikli bireylerde de hayatı tehdit eden sorunlar oluşturabilir. Bu çalışmada, hematopoetik kök hücre nakli yapılmış çocuklarda, kolistin ve seftazidim-avibaktam temelli tedavilerin etkilerini karşılaştırmayı amaçladık. Şubat 2019-Mayıs 2021 tarihleri arasında hematopoetik kök hücre nakli yapılan pediatrik hastaların tıbbi kayıtları retrospektif olarak değerlendirildi. Demografik bilgiler, altta yatan hastalıklar, hastanın hastaneye kabulü sırasındaki hastalığın şiddeti, seftazidim-avibaktam verilmesini gerektiren ilk vakanın ayrıntılı açıklaması, kolistin, seftazidim-avibaktam veya diğer antibakteriyel uygulamalarla ilgili veriler, ilaç tedavisi sonrası gözlenen klinik ve mikrobiyolojik yanıtlar ve ilaç yan etkilerini kaydetmek için önceden yapılan dırılmış bir vaka raporu formu kullanıldı. Hematopoetik kök hücre nakli yapılan 183 pediatrik hastada 89 bakteriyel enfeksiyon tespit edildi ve 57 hastada 72 gram-negatif mikroorganizma izole edildi. Bu hastaların 14 (%24.6)'ü, 18 (%25)'i çoklu ilaç dirençli gram-negatif mikroorganizmaya sahipti. Kolistin temelli tedavi alan yedi hastanın 5 (%71)'inde nefrotoksisite, 2 (%28.5)'inde hepatotoksisite gelişti. Seftazidim-avibaktam temelli tedavi grubunda 3 (%42.8) hastada nefrotoksisite, 2 hastada (%28.5)

Correspondence Address/Yazışma Adresi

Barış Malbora

Unit of Pediatric Stem Cell Transplant,
Yeni Yüzyıl Üniversitesi Gaziosmanpaşa Hospital,
İstanbul, Türkiye

E-mail: barismalbora@gmail.com

Received: 14.09.2024 Accepted: 10.05.2025

Available Online Date: 17.03.2026

This work is licensed under the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License.

Data Sharing Statement: The data that support the findings of this study are available from the corresponding author upon reasonable request.

©Copyright 2025 by Pediatric Infectious Diseases and Immunization Society. Available online at www.cocukenfeksiyon.org

hepatotoxicity occurred in 2 (28.5%) patients. Nephrotoxicity developed in 3 (42.8%) patients and hepatotoxicity in 2 (28.5%) patients in the ceftazidim-avibactam-based treatment group. Ten (71.4%) patients had clinical and microbiologic responses; however four patients died for various causes. In our study, we found that both colistin and ceftazidime-avibactam had similar survival rates in multidrug-resistant gram-negative infections in children who underwent hematopoietic stem cell transplantation. Organ toxicities observed in patients were mostly temporary. In addition, we observed that nephrotoxicity was less common in patients receiving ceftazidime-avibactam-based treatment. We think that colistin or ceftazidime-avibactam-based antibiotic combinations can be used successfully in pediatric patients with multidrug-resistant gram-negative infections.

Keywords: Multidrug-resistant gram-negative bacteria, ceftazidim-avibactam, colistin, hematopoietic stem cell transplantation, child

Introduction

Antimicrobial resistance is a life-threatening global problem for hematopoietic stem cell transplant (HSCT) patients. Multidrug resistant (MDR) gram-negative bacteria have serious effects on transplant patients, increasing the likelihood of bacterial colonization while increasing the risk of severe invasive infection, graft failure, and death (1,2). These microorganisms are resistant to almost all antibiotics, including carbapenems. Therefore, treatment options are limited to a few broad-spectrum antimicrobial drugs such as colistin (CST) (3,4). Unfortunately, acquired resistance to CST has emerged and become a global problem (5). Carbapenem-resistant *Enterobacteriaceae* (CRE) have recently emerged in the pediatric population and have led to outbreaks of infections that have caused an increase in mortality due to the greatly limited treatment options (6,7). Since there is virtually no safety and pharmacokinetic data on CST in these patient groups, these organisms behave as if they are pan-drug resistant (PDR). Recently, the combination of ceftazidime and avibactam has been approved for the treatment of CR-GNB (8). Various clinical studies using ceftazidime-avibactam (C/A) for CRE infection have shown that the overall response rate ranged from 45% to 74% (8). To date, no studies have evaluated the use of C/A in pediatric HSCT patients with Gram-negative infections. In this study, we aimed to compare the clinical outcomes and side effects of existing drugs in children with HSCT and CRS Gram-negative sepsis and to examine the effects and toxicities of CST-based and C/A-based antibacterial treatments.

Materials and Methods

Research Design

This study includes a retrospective observation of pediatric patients who underwent HSCT between February 2019 and May 2021 and were infected with Gram-negative microorganisms. The study was conducted at the Pediatric Bone Marrow Transplantation Unit of İstanbul Yeni Yüzyıl University Gaziosmanpaşa Hospital (EBMT CIC number: 0459).

hepatotoksosite gelişti. Toplamda 10 (%71.4) hastada klinik ve mikrobiyolojik yanıt elde edildi. Ancak dört hasta farklı nedenlerle hayatını kaybetti. Çalışmamızda, hematopoetik kök hücre nakli yapılmış çocuklarda çoklu ilaç dirençli gram-negatif enfeksiyonlarda hem kolistin hem de seftazidim-avibaktamın benzer sağkalım oranlarına sahip olduğunu saptadık. Hastalarda görülen organ toksisiteyi çoğunlukla geçiciydi. Buna ek olarak seftazidim-avibaktam temelli tedavi alan hastalarda nefrotoksitenin daha az görüldüğünü gözlemledik. Çoklu ilaç dirençli gram-negatif enfeksiyonlu pediyatrik hastalarda kolistin veya seftazidim-avibaktam tabanlı antibiyotik kombinasyonlarının başarılı bir şekilde kullanılabileceğini düşünmekteyiz.

Anahtar Kelimeler: Çoklu ilaç dirençli gram-negatif bakteriler, seftazidim-avibaktam, kolistin, hematopoetik kök hücre nakli, çocuk

Our center performs an average of 90 HSCTs per year. The database created examined all cases of gram-negative infections occurring between February 1, 2019, and May 31, 2020. Additionally, the clinical records of patients with MDR gram-negative infections were obtained. All demographic data, underlying primary diseases, transplantation procedures, MDR gram-negative infection episodes, and survival data were recorded. All MDR gram-negative infections that occurred after the start of the preparation regimen for bone marrow transplantation were analyzed. CST was preferred for the treatment of individuals infected with carbapenem-resistant and CST-sensitive bacteria. Only infectious disease specialists were permitted to prescribe C/A for the treatment of microbiologically confirmed MDR Gram-negative infections in critically ill patients. Both antibiotics were combined with other antibiotics for an additive effect.

Data Analysis

At least two researchers evaluated the medical, laboratory, and pharmacological data of eligible individuals. A case report form was used to record demographic information, comorbidities and primary diseases, disease severity at admission, a detailed description of the infection requiring C/A, data related to C/A, the dose and duration of CST or other antimicrobials, clinical and laboratory responses, side effects, and outcomes.

Definitions

Criteria from the United States Centers for Disease Control and Prevention were used to define hospital-acquired infections (9). Therefore, MDR refers to the absence of acquired susceptibility to at least one agent in three or more antimicrobial categories, while extremely drug resistant refers to the absence of acquired susceptibility to all agents in at least two or fewer antimicrobial categories (10). PDR indicates the absence of acquired susceptibility to all agents in all existing antimicrobial categories (10). If at least one strain of *Escherichia coli*, *Klebsiella aerogenes*, *Klebsiella oxytoca*, *Klebsiella pneumoniae*, or *Enterobacter* spp. is proven

to be resistant to the following antibiotics: 1) imipenem, 2) imipenem/rellebactam, 3) meropenem, 4) doripenem, 5) ertapenem, or 6) meropenem/vaborbactam, these are classified as “CRE” (10). 3. The degree of organ toxicity was analyzed based on the National Cancer Institute’s “Common Toxicity Criteria for Adverse Events” scales (11).

Evaluation of the results

The first negative culture obtained after initiating CST or C/A-based therapy was used to determine the timing of the microbiological response. Improvement/remission of the main symptoms was defined as the clinical response. The following outcomes were evaluated within 15 days after starting CST or C/A: If all symptoms had resolved and no further antimicrobial treatment was required, it was considered a “cure, complete recovery”; if symptoms had improved but antimicrobial treatment was continuing, it was considered “improvement”; and if there was worsening or death, it was considered “failure”. Additionally, long-term outcomes within 30 days after starting CST or C/A-based treatment were examined using the 15-day protocol, but relapse was also considered.

Safety Assessment

To evaluate patients during CST and C/A treatment, the World Health Organization International Drug Monitoring Center adverse reaction standard criteria were used (12).

Microbiological Tests

Both clinical and surveillance data were used in this study. The BACT-Alert system (bioMerieux®, Marcy l’Etoile, France) was used for blood cultures. The VITEK-2 automated system (bioMerieux®) was used for diagnosis and antimicrobial

susceptibility testing. C/A susceptibility was determined by E-test according to the following susceptibility breakpoints: zone diameter 13 mm (C/A disk content: 10-4 g) and minimum inhibitory concentration (MIC) 8 g/mL. The reference medium microdilution technique was used to determine CST susceptibility (13).

Results

Between February 2019 and May 2021, a total of 183 pediatric patients underwent HSCT. During this period, 89 bacterial agents were detected in 72 patients (39.3%) in different regions. A total of 72 (80.9%) gram-negative infections were detected in 52 (72.2%) of these patients. A total of 18 (25%) MDR gram-negative infections were detected in 14 (27%) of these 52 patients (Figure 1). All these microbial agents were resistant to carbapenems, and the MIC of all microorganisms was at least 16, while the ertapenem MIC value was at least 8. The choice of treatment for patients was determined based on the results of the antibiotic susceptibility test. All patients continued to show signs of sepsis despite empirical treatment. Seven patients received CST-based combination therapy. The remaining patients received C/A-based combination therapy. The demographic, microbiological, and clinical characteristics of these patients are shown in Table 1 and Table 2.

Colistin-based therapy

The mean time between microbial agent growth in culture and initiation of CST was 2.6 days (range 1-4 days). CST was administered at the recommended dose of 5 mg/kg twice daily based on antibiotic susceptibility and was used for an average of 14 days (range 5 to 22 days). All patients received combination therapy with CST. The microorganism could

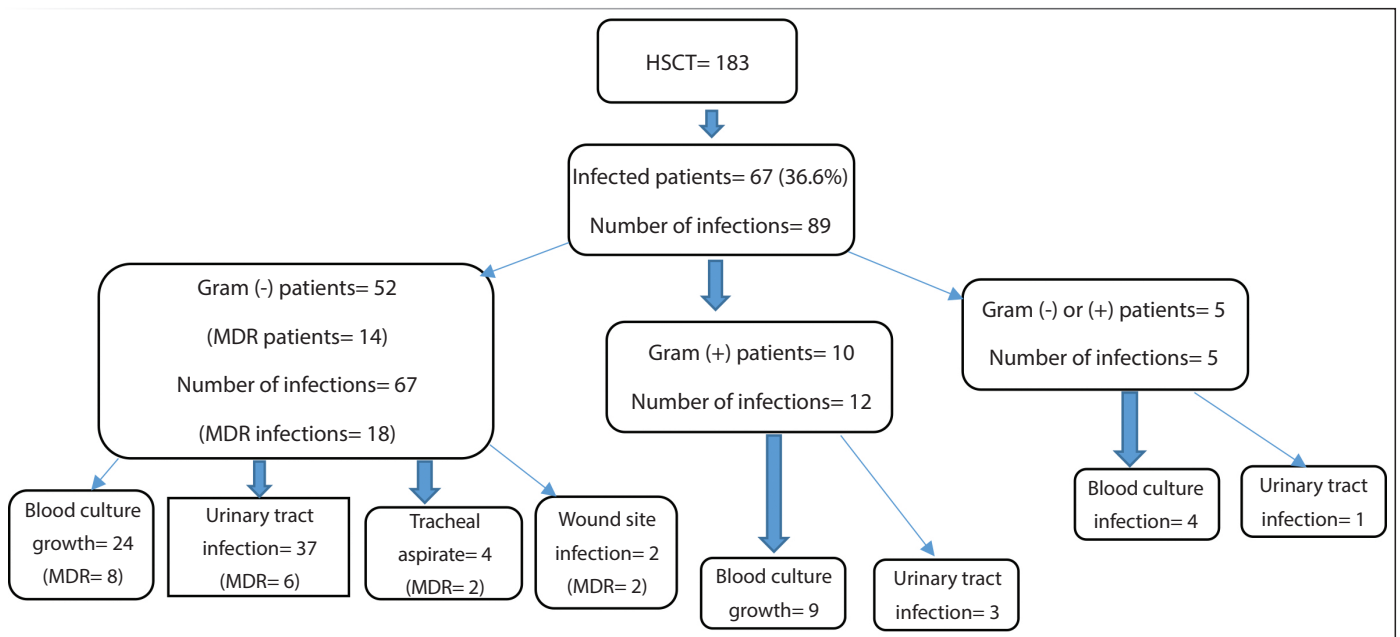


Figure 1. Bacterial distribution pattern in patients undergoing hematopoietic stem cell transplantation.

Table 1. Demographic, microbiological, and clinical characteristics of the patients using colistin

Patient #	1	2	3	4	5	6	7
Age/Sex	14 mo/F	9 y/M	15 y/M	6 y/F	11 y/F	13 y/F	15 y/M
Patient's country of origin	Türkiye	Tunisia	Iraq	Iraq	Iraq	Türkiye	Iraq
Disease	HLH	ALL	SAA	PNH-AAA	FAA	TM	SAA
HSCT type/Source	Sibling/BM	Haplo/PDSC	Sibling/BM	Sibling/BM	SiblingBM	Non-relative/PDSC	Non-relative/ PDSC
Microbial agent	<i>P. aeruginosa</i>	<i>P. aeruginosa</i>	<i>Elizabethkingia meningoseptica</i>	<i>E. coli</i>	<i>E. coli</i>	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>
Site of the microorganism	Wound site	Urinary tract infection	Blood culture growth+urinary tract infection	Urinary tract infection	Blood culture growth	Blood culture growth+urinary tract infection	Blood culture growth+wound site growth
Post-transplant (day)	20	3	86	8	2	52	13
Neutropenia during infection	Yes	Yes	No	Yes	Yes	No	No
Accompanying problems	No	No	TMA+skin/ liverGvHD+PRES	VOD+ATN	No	GI GvHD+VOD+PRES	Liver GvHD
ICU admission	No	No	No	No	No	Yes	No
Intubation	No	No	No	No	No	Yes	No
Inotropic support	No	No	No	No	No	Yes	No
Prior antibiotic exposure	PIP-TAZ	MEM	Yok	MEM+AMK	SEF	SEF+FOS+AMK	MEM+AMK
Time elapsed between the onset of infection and CST initiation (day)	3	2	4	2	4	2	1
CST duration (day)	14	5	20	15	10	12	22
Other antibiotics with CST	PIP-TAZ	MEM	TGC+MEM	AMK	MEM	MEM	TGC
Microorganism eradication time (day)	12	4	No eradication	4	6	No eradication	17
Side effects	No	Nephrotoxicity/ Hepatotoxicity	Nephrotoxicity/ Hepatotoxicity	Nephrotoxicity	No	Nephrotoxicity	Nephrotoxicity
Outcomes/Reasons	Alive	Alive	Exitus/sepsis	Alive	Alive	Sexitus/sepsis and GI GvHD	Alive

F: Female, M: Male, y: Age, HLH: Hemophagocytic lymphohistiocytosis, ALL: Acute lymphoblastic leukemia, AAA: Severe aplastic anemia, PNH: Paroxysmal nocturnal hemoglobinuria FAA: Fanconi aplastic anemia, TM: Thalassemia major, PHSC: Peripheral-derived stem cell, BM: Bone marrow, *P. aeruginosa*: *Pseudomonas aeruginosa*, *E. coli*: *Escherichia coli*, *K. pneumoniae*: *Klebsiella pneumoniae*, VOD: Venocclusive disease, ATN: Acute tubular necrosis, TMA: Thrombotic microangiopathy, GvHD: Graft versus host disease, PRES: Posterior reversible encephalopathy syndrome, PIP-TAZ: Pirasilin-tazobactam, MEM: Meropenem, AMK: Amikacin, SEF: Ceftazidime-avibactam, FOS: Foscarnet, TGC: Tigecycline.

not be eradicated in two patients (patients #3 and #6). These patients died of sepsis on the 15th and 12th days of CST-based therapy, respectively. In the remaining five patients, the microorganisms were eradicated within an average of 8 days (range 4-14 days) after starting CST. Nephrotoxicity developed in 5 of the 7 (71%) patients receiving CST-based therapy, while hepatotoxicity developed in 2 (28.5%) patients. Side effects were reversible in all patients except one (patient #3) (Table 1).

Ceftazidime-avibactam-based therapy

The mean time between microbial agent growth in culture and initiation of C/A was 4.8 days (range 1-10 days). C/A was added to the existing antibiotic combination after a mean of

eight days (range 3-14 days) following administration of the recommended dose of 62.5 mg/kg three times daily based on antibiotic susceptibility. All patients received combination therapy with C/A. The microbial agent was eradicated within an average of 4.3 days (range 1-11 days) after starting C/A therapy. Subsequently, a clinical and microbiological response was obtained in five of the patients. Patients #10 and #14 died on the 13th and 12th day of C/A treatment, respectively, due to excessive bleeding and respiratory failure (Table 2). Nephrotoxicity developed in 3 (42.8%) patients and hepatotoxicity in 2 (28.5%) patients in the C/A-based treatment group. The three patients who developed nephrotoxicity were

Table 2. Demographic, microbiological, and clinical characteristics of the patients using ceftazidime-avibactam

Patient #	8	9	10	11	12	13	14
Age/Sex	26 mo/M	89 mo/M	18 y/M	35 mo/M	48 mo/F	25 mo/F	93 mo/M
Patient's country of origin	Türkiye	Russia	Tunusia	Kosova	Ukraine	Türkiye/Arab origin	Ukraine
Disease	ALL	AML	ALL	NB	NB	TM	DK
HSCT type/Source	Non-relative/PDSC	Haplo/PDSC	Non-relative/PDSC	Autologous/PDSC	Non-relative/PDSC	Relative/BM	Non-relative/PDSC
Microbial agent	<i>K. pneumoniae</i>	<i>P. aeruginosa</i>	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	<i>E. coli</i> and <i>K. pneumoniae</i>	<i>K. pneumoniae</i>	<i>Chryseobacterium indolegenes</i>
Site of the microorganism	Blood culture growth	Tracheal aspirate	Blood culture growth	Urinary tract infection	Blood culture growth+tracheal aspirate	Blood culture growth	Urinary tract infection
Post-transplant (gün)	+20	+86	+8	+6	+36	-15	+82
Neutropenia during infection	No	No	Yes	Yes	No	Pre-transplantation	No
Accompanying problems	Respiratory failure	Cerebral toxoplasmosis	Respirator failure primary graft failure	No	Respiratory failure	Respiratory failure	MDR+CMV reactivation+secondary graft failure
ICU admission	No	Yes	Yes	Yes	Yes	Yes	Yes
Intubation	No	Yes	Yes	Yes	Yes	Yes	Yes
Inotropic support	No	Yes	Yes	Yes	Yes	Yes	Yes
Prior antibiotic exposure	CST+MEM	MEM+AMK	CST+AMK	MEM	MEM+AMK	MEM+AMK	CST
Time elapsed between the onset of infection and C/A initiation (day)	4	6	3	3	10	3	1
C/A duration (day)	8	17	13	7	12	10	12
Other antibiotics with C/A	TGC+FOS	MEM	CST+AMK	MEM	CST	CST+AMK	MEM
Microorganism eradication time (day)	8	13	14	4	14	5	N/A
Side effects	No	No	Nephrotoxicity/Hepatotoxicity	Nephrotoxicity	Nephrotoxicity	No	Nephrotoxicity/Hepatotoxicity
Outcomes/Reasons	Alive	Alive	Exitus/sepsis	Alive	Alive	Alive	Exitus/sepsis and massive GI bleeding

F: Female, M: Male, y: Age, HLH: Hemophagocytic lymphohistiocytosis, ALL: Acute lymphoblastic leukemia, AAA: Severe aplastic anemia, PNH: Paroxysmal nocturnal hemoglobinuria FAA: Fanconi aplastic anemia, TM: Thalassemia major, PHSC: Peripheral-derived stem cell, BM: Bone marrow, *P. aeruginosa*: *Pseudomonas aeruginosa*, *E. coli*: *Escherichia coli*, *K. pneumoniae*: *Klebsiella pneumonia*, VOD: Venooclusive disease, ATN: Acute tubular necrosis, TMA: Thrombotic microangiopathy, GvHD: Graft versus host disease, PRES: Posterior reversible encephalopathy syndrome, MDR: Multidrug resistant, CMV: Cytomegalovirus PIP-TAZ: Piperacillin-tazobactam, CST: Colistin, MEM: Meropenem, AMK: Amikacin, SEF: Ceftazidime-avibactam, FOS: Foscarnet, TGC: Tigecycline, N/A: Not available.

also receiving other drugs with nephrotoxic properties, such as amikacin and CST. There were no treatment interruptions due to side effects. Side effects were reversible except in one patient who died due to sepsis and massive gastrointestinal bleeding (patient #14) (Table 1).

Discussion

Infections caused by MDR gram-negative bacteria are becoming increasingly common in HSCT recipients. Invasive MDR gram-negative infections are associated with high mortality rates in transplant recipients due to limited treatment

options and antimicrobial toxicity (14). At our center, a total of 183 pediatric patients underwent HSCT between February 2019 and May 2021. During this period, 89 bacterial agents were detected in 72 patients (39.3%) in different regions. A total of 72 (80.9%) gram-negative infections were detected in 52 (72.2%) of these patients. A total of 18 (25%) MDR gram-negative infections were present in 14 (27%) of these 52 patients (Figure 1). All of these microbial agents were resistant to carbapenems, and the MIC of all microorganisms was at least 16, while the MIC value of ertapenem was at least 8. The choice of treatment for patients was determined based on the results of the antibiotic susceptibility test. All patients showed persistent septic symptoms despite empirical treatment. Seven patients received CST-based combination therapy. The remaining patients received C/A-based combination therapy.

In children, the incidence of infections caused by MDR bacteria has increased in line with that seen in adult populations. The mortality rate for individuals with carbapenem-resistant-*K. pneumoniae* ranges from 22% to 45%. This rate is higher in patients with concurrent infections or comorbidities (15). According to data from the last decade, the prevalence of carbapenem resistance in children in the United States increased from 0% to 0.47% among *Enterobacterales* isolates between 2000 and 2011, *Pseudomonas aeruginosa* isolates increased from 9.4% to 20% between 1999 and 2012, and among *Acinetobacter baumannii* isolates, it increased from 0.6% to 6.1% between 1999 and 2012 (16-18). Between 2012 and 2013, a multicenter study in Italy found a twofold increase in CRE colonization and a fourfold increase in the incidence of CRE bloodstream infections (19).

A large-scale study conducted in the United States between 2012 and 2013 found that only 0.6% of children under the age of 18 were infected with CRE (20). CRE is frequently isolated in pediatric units when it is highly endemic. In non-endemic regions, it is usually detected in adult intensive care units and oncology services (21-23). Therefore, internationally published pediatric data (United Kingdom, Spain, Italy, and United States) have been consistent with sporadic outbreaks of CRE infections (24-26).

Pediatric CRE infections have been shown to increase the risk of death by 6 to 11 times compared to non-CRE infections (27,28). On the other hand, there are also many studies that have found no worse outcomes in pediatric patients with CRE infections (29). Depending on the source of infection, the primary disease, and age, mortality rates in children with CRE infections range from 8% to 52% (in one small case series, the rate was 100%) (19,26-33). In our study, the mortality rate in MDR gram-negative infections was 28% (4/14). One of the patients who died in the CST group had severe aplastic anemia and severe concomitant diseases [transplant-related microangiopathy, grade III graft versus host disease (GvHD),

and posterior reversible encephalopathy syndrome (PRES)]. The other was a patient diagnosed with thalassemia major who had severe gastrointestinal GvHD and PRES. Both patients were receiving multiple immunosuppressive drugs. One of the patients who died in the C/A group was a patient diagnosed with acute lymphoblastic leukemia (ALL) who had not yet achieved neutrophil engraftment. The other was a patient diagnosed with congenital dyskeratosis who experienced secondary graft failure due to MDR cytomegalovirus infection and massive gastrointestinal bleeding.

A recently published study identified several important risk factors for mortality in 50 children with hospital-acquired CRE bloodstream infections. These included admission to a pediatric intensive care unit, intubation, and inotropic support (31). In this review, a meropenem MIC value >8 mg/L for the isolate significantly increased the likelihood of death [Overall rate, 13.9 (95% confidence interval, 1.5 to 125.6); $p=0.008$]. All of our patients (except patient #1) were admitted to the intensive care unit, received inotropic support, and required mechanical ventilation.

Risk factors for CRE infection or colonization in children are comparable to those defined for adults. Most had underlying comorbidities or a history of intensive care unit admission (34-36). Prior antibiotic exposure (usually long-term broad-spectrum antibiotics), medical device use (especially mechanical ventilation), intensive care unit stay, previous surgical interventions, and prolonged hospital stay are risk factors for CRE colonization or infection in children (26,27,29,37-41). A recent study found that only carbapenem exposure was significantly associated with CRE infections in children (38). Other pediatric studies have shown that past carbapenem use is a risk factor for CRE infection (27,29,37,38,40,41). However, the association with non-carbapenem antibiotic use was less clear (29,39,40). With one exception (patient #6), all patients had a history of broad-spectrum antibiotic use. All of these patients were being treated for malignancy, immunodeficiency, or bone marrow failure. All patients had a catheter and were hospitalized for a long time because they were undergoing transplantation. Patient #11 had a history of previous surgery. Four patients (patient #6, #8, #12, and #14) were infected with MDR gram-negative microorganisms while being monitored in the intensive care unit.

Individuals from regions endemic for carbapenem-resistant enterococci can carry CRE to medical centers in non-endemic countries, which may require special precautions such as colonization screening and antibiotic treatment (21-23,37,39). Interestingly, all patients except four (patient #1, #6, #8, and #13) were from outside Türkiye (Tables 1 and 2).

The safety and pharmacokinetics of meropenem in children have been studied in several investigations (42,43). When higher doses of meropenem are administered via

prolonged infusion in adults, the likelihood of achieving the pharmacodynamic target is higher for isolates with an MIC value of 8 mg/L (44). This strategy has been applied to pediatric patients with CRE infections (45). In pediatric population modeling, methods involving prolonged infusion and higher doses (40 mg/kg/8 hours per body weight) increased the likelihood of achieving meropenem bactericidal targets (46). In two pediatric studies, infections with isolates having a meropenem MIC value greater than 8 mg/L were associated with increased mortality, and all patients associated with isolates having a meropenem MIC value greater than 32 mg/L died (31). In this study, children infected with isolates with MIC values greater than 8 mg/L died at a higher rate than children receiving meropenem for isolates with MIC values less than 8 mg/L (100% vs. 45.5%). Furthermore, significant concerns have been raised regarding the likelihood of achieving the target MIC value of 4 to 8 mg/L for gram-negative pathogens in critically ill pediatric populations (46). Zheng et al. demonstrated that combining C/A with an antibiotic that is in vitro non-susceptible, such as carbapenems, tigecycline, and fosfomycin, could significantly reduce the 30-day mortality rate in critically ill patients with CR-K. *pneumoniae* infection (47). Due to differences between zinc concentrations in routine media and those at infection sites, carbapenem resistance in *Enterobacteriaceae* may be misdiagnosed using antimicrobial susceptibility testing techniques (48,49). These findings suggest that certain CREs examined in in vitro antimicrobial susceptibility tests may be in vivo susceptible. In clinical practice, polymyxins (polymyxin B and CST) have been considered as last-resort antibiotics for managing CRKP infections prior to C/A (50,51). However, there are no clinical studies evaluating a combination of C/A and polymyxins. Patients in both the CST and C/A groups were treated with different antibiotic combinations. Additionally, four patients in the CST group (patient #2, #3, #5, #6) and three patients in the C/A group (patient #9, #11, and #14) received concomitant meropenem therapy. We administered meropenem as an intravenous infusion at a dose of 40 mg/kg every eight hours. Other patients were treated with various combinations such as piperacillin-tazobactam, amikacin, tigecycline, and fosfomycin (Table 1,2).

C/A is approved in the European Union and the United States for the treatment of other infections caused by gram-negative organisms that can be treated with limited success in adults, including complicated urinary tract infections, pyelonephritis, complicated intra-abdominal infections, hospital-acquired pneumonia, and ventilator-associated pneumonia (52-54).

In recent years, MDR gram-negative infections have become a serious public health problem. The treatment of these infections is particularly difficult in

immunocompromised children who have undergone HSCT and is associated with high mortality rates. Assessing risk factors for acquiring MDR gram-negative infections based on local epidemiology may enable personalized empirical broad-spectrum antibiotic therapy. Clinical data for the treatment of MDR gram-negative infections are still limited and mostly based on observational studies. This is even more limited in these children. Antibiotic stewardship programs remain an important component in preserving the current effectiveness of antibiotics with a sensitive approach to antimicrobial treatment. New antibiotics can provide very successful treatments. However, the lack of pediatric studies leads to off-label use in this group.

This study has some limitations, such as being a retrospective cohort study with a small sample size. Prospective or randomized controlled trials with more participants should be designed for future research. Second, mutations responsible for antibiotic resistance mechanisms could not be analyzed in our study.

In conclusion, CST appears to be a good option in children with MDR gram-negative infections who have undergone HSCT. However, if these microorganisms are also resistant to CST, the use of C/A may be life-saving in immunocompromised patients.

Peer-review: Externally peer-reviewed.

Author Contributions: Concept - BM, DBT; Design - HS; Supervision - DBT; Resource - ZD; Data Collection and/or processing - BM, HS; Analysis and/or interpretation - DE, BM; Literature search - AAA, BM; Writing - BM, DBT; Critical review - BM, DBT, AAA.

Conflict of Interest: All authors declare that they have no conflict of interest.

Financial Disclosure: The authors declared that this study has received no financial support.

References

1. Bartoletti M, Giannella M, Tedeschi S, Viale P. Multidrug-resistant bacterial infections in solid organ transplant candidates and recipients. *Infect Dis Clin North Am* 2018;32(3):551-80. <https://doi.org/10.1016/j.idc.2018.04.004>
2. Theuretzbacher U. Global antimicrobial resistance in gram-negative pathogens and clinical need. *Curr Opin Microbiol* 2017;39:106-12. <https://doi.org/10.1016/j.mib.2017.10.028>
3. Magiorakos AP, Srinivasan A, Carey RB, Falagas ME, Giske CG, Harbarth S, et al. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect* 2012;18:268-81. <https://doi.org/10.1111/j.1469-0691.2011.03570.x>
4. Tumbarello M, Losito AR, Giamarellou H. Optimizing therapy in carbapenem-resistant *Enterobacteriaceae* infections. *Curr Opin Infect Dis* 2018;31:566-77. <https://doi.org/10.1097/QCO.0000000000000493>

5. Karaiskos I, Souli M, Galani I, Giamarellou H. Colistin: still a life saver for the 21st century? *Expert Opin Drug Metab Toxicol* 2017;13:59-71. <https://doi.org/10.1080/17425255.2017.1230200>
6. Logan LK. Carbapenem-resistant Enterobacteriaceae: an emerging problem in children. *Clin Infect Dis* 2012;55:852-9. <https://doi.org/10.1093/cid/cis543>
7. Trecarichi EM, Pagano L, Candoni A, Pastore D, Cattaneo C, Fanci R, et al. Current epidemiology and antimicrobial resistance data for bacterial kan yolu infections in patients with hematologic malignancies: an Italian multicentre prospective survey. *Clin Microbiol Infect* 2015;21(4):337-43. <https://doi.org/10.1016/j.cmi.2014.11.022>
8. Wright H, Bonomo RA, Paterson DL. New agents for the treatment of infections with Gram-negative bacteria: restoring the miracle or false dawn? *Clin Microbiol Infect* 2017;23(10):704-12. <https://doi.org/10.1016/j.cmi.2017.09.001>
9. Magiorakos AP, Carey SRB, Carmeli Y, Falagas ME, Giske CG, Harbarth S, et al. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect* 2012;18(3):268-81. <https://doi.org/10.1111/j.1469-0691.2011.03570.x>
10. Centers for Disease Control and Prevention (CDC). Antimicrobial resistant phenotype definitions. Available from: <https://www.cdc.gov/ncezid/updates/nhsn-maintenance.html> (Accessed date: 30.11.2024).
11. National Cancer Institute. Common Terminology Criteria for Adverse Events v4.0. NIH publication 09-7473. May 29 2009, NCI, NIH, DHHS.
12. Edwards IR, Biriell C. Harmonisation in pharmacovigilance. *Drug Saf* 1994;10:93-102. <https://doi.org/10.2165/00002018-199410020-00001>
13. The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters, version 9.0, 2019. Available from: http://www.eucast.org/clinical_breakpoints/
14. Cairns KA, Hall V, Martin GE, Griffin DWJ, Stewart JD, Khan SF, et al. Treatment of invasive IMP-4 Enterobacter cloacae infection in transplant recipients using ceftazidime/avibactam with aztreonam: A case series and literature review. *Transpl Infect Dis* 2021;23(2):e13510. <https://doi.org/10.1111/tid.13510>
15. Munoz-Price LS, Poirel L, Bonomo RA, Schwaber MJ, Daikos GL, Cormican M, et al. Clinical epidemiology of the global expansion of Klebsiella pneumoniae carbapenemases. *Lancet Infect Dis* 2013;13(9):785-96. [https://doi.org/10.1016/S1473-3099\(13\)70190-7](https://doi.org/10.1016/S1473-3099(13)70190-7)
16. Logan LK, Renschler JP, Gandra S, Weinstein RA, Laxminarayan R, Centers for Disease Control and Prevention Epicenters Program. Carbapenem-resistant Enterobacteriaceae in children, United States, 1999-2012. *Emerg Infect Dis* 2015;21:2014-21. <https://doi.org/10.3201/eid2111.150548>
17. Logan LK, Gandra S, Mandal S, Klein EY, Levinson J, Weinstein RA, et al. Prevention Epicenters Program, U.S. Centers for Disease Control and Prevention. Multidrug- and carbapenem-resistant Pseudomonas aeruginosa in children, United States, 1999-2012. *J Pediatric Infect Dis Soc* 2016;6:352-9. <https://doi.org/10.1093/jpids/piw064>
18. Logan LK, Gandra S, Trett A, Weinstein RA, Laxminarayan R. Acinetobacter baumannii resistance trends in children in the United States, 1999-2012. *J Pediatric Infect Dis Soc* 2019;8:136-42. <https://doi.org/10.1093/jpids/piy018>
19. Caselli D, Cesaro S, Fagioli F, Carraro F, Ziino O, Zanazzo G, et al. Incidence of colonization and kan yolu infection with carbapenem-resistant Enterobacteriaceae in children receiving antineoplastic chemotherapy in Italy. *Infect Dis (Lond)* 2016;48:152-5. <https://doi.org/10.3109/23744235.2015.1087647>
20. Guh AY, Bulens SN, Mu Y, Jacob JT, Reno J, Scott J, et al. Epidemiology of carbapenem-resistant Enterobacteriaceae in 7 US communities, 2012-2013. *JAMA* 2015;314:1479-87. <https://doi.org/10.1001/jama.2015.12480>
21. Jajoo M, Manchanda V, Chaurasia S, Sankar MJ, Gautam H, Agarwal R, et al. Alarming rates of antimicrobial resistance and fungal sepsis in outborn neonates in North India. *PLoS One* 2018;13:e0180705. <https://doi.org/10.1371/journal.pone.0180705>
22. Sekar R, Mythreyee M, Srivani S, Sivakumaran D, Lallitha S, Saranya S. Carbapenem-resistant Enterobacteriaceae in pediatric infections in rural southern India. *Indian Pediatr* 2017;54:1021-4. <https://doi.org/10.1007/s13312-017-1204-1>
23. Alp E, Perçin D, Colakoglu S, Durmaz S, Kurkcu CA, Ekincioglu P, et al. Molecular characterization of carbapenem-resistant Klebsiella pneumoniae in a tertiary university hospital in Turkey. *J Hosp Infect* 2013;84:178-80. <https://doi.org/10.1016/j.jhin.2013.03.002>
24. Herruzo R, Ruiz G, Perez-Blanco V, Gallego S, Mora E, Vizcaino MJ, et al. Bla-OXA48 gene microorganisms outbreak, in a tertiary children's hospital, over 3 years (2012-2014): Case report. *Medicine (Baltimore)* 2017;96:e7665. <https://doi.org/10.1097/MD.00000000000007665>
25. Alvares PA, Arnoni MV, da Silva CB, Sáfadi MAP, Mimica MJ. Hospital-acquired infections in children: A Latin American Tertiary teaching hospital 5-year experience. *Pediatr Infect Dis J* 2019;38(1):e12-e14. <https://doi.org/10.1097/INF.0000000000002046>
26. Montagnani C, Prato M, Scolfaro C, Colombo S, Esposito S, Tagliabue C, et al. Carbapenem resistant Enterobacteriaceae infections in children. *Pediatr Infect Dis J* 2016;35:862-8. <https://doi.org/10.1097/INF.0000000000001188>
27. Alvares PA, Arnoni MV, da Silva CB, Sáfadi MAP, Mimica MJ. Carbapenem-resistant Gram-negative infections in critically ill children: outcome and risk factors in a tertiary teaching hospital in South America. *J Hosp Infect* 2019;101:188-9. <https://doi.org/10.1016/j.jhin.2018.10.001>
28. Chiotos K, Tamma PD, Flett KB, Karandikar MV, Nemati K, Bilker WB, et al. Increased 30-day mortality associated with carbapenem-resistant Enterobacteriaceae in children. *Open Forum Infect Dis* 2018;5:222. <https://doi.org/10.1093/ofid/ofy222>
29. Logan LK, Nguyen DC, Scaggs Huang FA, Qureshi NK, Charnot-Katsikas A, Bartlett AH, et al. A multi-centered case-case control study of factors associated with Klebsiella pneumoniae carbapenemase-producing Enterobacteriaceae infections in children and young adults. *Pediatr Infect Dis J* 2019;38:490-5. <https://doi.org/10.1097/INF.0000000000002176>
30. Dong F, Zhang Y, Yao K, Lu J, Guo L, Lyu S, et al. Epidemiology of carbapenem-resistant Klebsiella pneumonia infections in a Chinese children's hospital: predominance of New Delhi metallo- β -lactamase-1. *Microb Drug Resist* 2018;24:154-60. <https://doi.org/10.1089/mdr.2017.0031>
31. Nabarro LEB, Shankar C, Pragasam AK, Mathew G, Jeyaseelan V, Veer-araghavan B, et al. Clinical and bacterial risk factors for mortality in children with carbapenem-resistant Enterobacteriaceae infections in India. *Pediatr Infect Dis J* 2017;36:161-6. <https://doi.org/10.1097/INF.0000000000001499>
32. Díaz A, Ortiz DC, Trujillo M, Garcés C, Jaimes F, Restrepo AV. Clinical characteristics of carbapenem-resistant Klebsiella pneumoniae infections in ill and colonized children in Colombia. *Pediatr Infect Dis J* 2016;35:237-41. <https://doi.org/10.1097/INF.0000000000000987>
33. Taoufik L, Amrani Hanchi A, Fatiha B, Nissrine S, Mrabih Rabou MF, Nabila S. Emergence of OXA-48 carbapenemase producing Klebsiella pneumoniae in a neonatal intensive care unit in Marrakech, Morocco. *Clin Med Insights Pediatr* 2019;13:1179556519834524. <https://doi.org/10.1177/1179556519834524>

34. Pannaraj PS, Bard JD, Cerini C, Weissman SJ. Pediatric carbapenem resistant Enterobacteriaceae in Los Angeles, California, a high prevalence region in the United States. *Pediatr Infect Dis J* 2015;34:11-6. <https://doi.org/10.1097/INF.0000000000000471>
35. Maltezou HC, Kontopidou F, Katerelos P, Daikos G, Roilides E, Theodoridou M. Infections caused by carbapenem-resistant Gram negative pathogens in hospitalized children. *Pediatr Infect Dis J* 2013;32:151-4. <https://doi.org/10.1097/INF.0b013e3182804b49>
36. Little ML, Qin X, Zerr DM, Weissman SJ. Molecular diversity in mechanisms of carbapenem resistance in paediatric Enterobacteriaceae. *Int J Antimicrob Agents* 2012;39:52-7. <https://doi.org/10.1016/j.ijantimicag.2011.09.014>
37. Nour I, Eldegl HE, Nasef N, Shouman B, Abdel-Hady H, Shabaan AE. Risk factors and clinical outcomes for carbapenem-resistant Gram-negative late-onset sepsis in a neonatal intensive care unit. *J Hosp Infect* 2017;97:52-8. <https://doi.org/10.1016/j.jhin.2017.05.025>
38. Sahbudak Bal Z, Bekmezci N, Soylu M, Sen S, Avcu G, Aydemir S, et al. The prospective evaluation of risk factors and clinical influence of carbapenem resistance in children with gram-negative bacteria infection. *Am J Infect Control* 2018;46:147-53. <https://doi.org/10.1016/j.ajic.2017.08.013>
39. Chiotos K, Tamma PD, Flett KB, Naumann M, Karandikar MV, Bilker WB, et al. Multicenter study of the risk factors for colonization or infection with carbapenem-resistant Enterobacteriaceae in children. *Antimicrob Agents Chemother* 2017;61:e01440-17. <https://doi.org/10.1128/AAC.01440-17>
40. Ozsurekci Y, Aykac K, Cengiz AB, Basaranoglu ST, Sancak B, Karahan S, et al. Kan yolu infections in children caused by carbapenem-resistant versus carbapenem-susceptible gram-negative microorganisms: risk factors and outcome. *Diagn Microbiol Infect Dis* 2017;87:359-64. <https://doi.org/10.1016/j.diagmicrobio.2016.12.013>
41. Ulu-Kilic A, Alp E, Percin D, Cevahir F, Altay-Kürkçü C, Ozturk AA. Risk factors for carbapenem resistant Klebsiella pneumoniae rectal colonization in pediatric units. *J Infect Dev Ctries* 2014;8:1361-4. <https://doi.org/10.3855/jidc.4593>
42. Cohen-Wolkowicz M, Poindexter B, Bidegain M, Weitkamp J-H, Scheinonka RL, Randolph DA, et al, Meropenem Study Team. 2012. Safety and effectiveness of meropenem in infants with suspected or complicated intra-abdominal infections. *Clin Infect Dis* 2012;55:1495-502. <https://doi.org/10.1093/cid/cis758>
43. Parker EM, Hutchison M, Blumer JL. The pharmacokinetics of meropenem in infants and children: a population analysis. *J Antimicrob Chemother* 1995;36:63-71. https://doi.org/10.1093/jac/36.suppl_A.63
44. Kuti JL, Dandekar PK, Nightingale CH, Nicolau DP. Use of Monte Carlo simulation to design an optimized pharmacodynamic dosing strategy for meropenem. *J Clin Pharmacol* 2033;43:1116-23. <https://doi.org/10.1177/0091270003257225>
45. Hsu AJ, Tamma PD. Treatment of multidrug-resistant Gram negative infections in children. *Clin Infect Dis* 2014;58:1439-48. <https://doi.org/10.1093/cid/ciu069>
46. Cies JJ, Moore WS, Enache A, Chopra A, Chopra A. Population pharmacokinetics and pharmacodynamic target attainment of meropenem in critically ill young children. *J Pediatr Pharmacol Ther* 2017;22:276-85. <https://doi.org/10.5863/1551-6776-22.4.276>
47. Zheng G, Zhang J, Wang B, Cai J, Wang L, Hou K, et al. Ceftazidim-avibactam in combination with in vitro non-susceptible antimicrobials versus seftazidim-avibaktam in monotherapy in critically ill patients with carbapenem-resistant Klebsiella pneumoniae infection: A retrospective cohort study. *Infect Dis Ther* 2021;10(3):1699-713. <https://doi.org/10.1007/s40121-021-00479-7>
48. Asempa TE, Abdelraouf K, Nicolau DP. Metallo-blactamase resistance in Enterobacteriaceae is an artefact of currently utilized antimicrobial susceptibility testing methods. *J Antimicrob Chemother* 2020;75(4):997-1005. <https://doi.org/10.1093/jac/dkz532>
49. Abdelraouf K, Reyes S, Nicolau DP. The paradoxical in vivo activity of b-lactams against metallo-b-lactamase-producing Enterobacteriales is not restricted to carbapenems. *J Antimicrob Chemother* 2021;76(3):684-91. <https://doi.org/10.1093/jac/dkaa467>
50. Karaiskos I, Lagou S, Pontikis K, Rapti V, Poulakou G. The, "old" and the "new" antibiotics for MDR gram-negative pathogens: for whom, when, and how. *Front Public Health* 2019;7:151. <https://doi.org/10.3389/fpubh.2019.00151>
51. Liang Q, Huang M, Xu Z. Early use of polymyxin B reduces the mortality of carbapenem-resistant Klebsiella pneumoniae infection. *Braz J Infect Dis* 2019;23(1):60-5. <https://doi.org/10.1016/j.bjid.2018.12.004>
52. Rodriguez BA, Giroto JE, Nicolau DP. Ceftazidime/avibactam and ceftolozane/tazobactam: novel therapy for multidrug resistant Gram negative infections in children. *Curr Pediatr Rev* 2018;14:97-109. <https://doi.org/10.2174/1573396314666180308150908>
53. European Medicines Agency. Zavicefta: summary of product characteristics, 2018. Available from: <http://www.ema.europa.eu> (Accessed date: 16.03.2018).
54. Food and Drug Administration. Avycaz (ceftazidime and avibactam) for injection, for intravenous use: US prescribing information, 2018. Available from: <https://www.accessdata.fda.gov> (Erişim tarih: 16.03.2018).